

Molecular hallmarks of lung cellular senescence in the bleomycin-induced and spirometry-confirmed mouse model of IPF

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Background & Aim

There is increasing evidence for senescent cells to play a contributory role in the pathogenesis of idiopathic pulmonary fibrosis (IPF).

This study aimed to characterize molecular markers of pulmonary senescence across the disease spectrum in the bleomycin-induced (BLEO) and spirometry-confirmed mouse model of IPF.

Methods

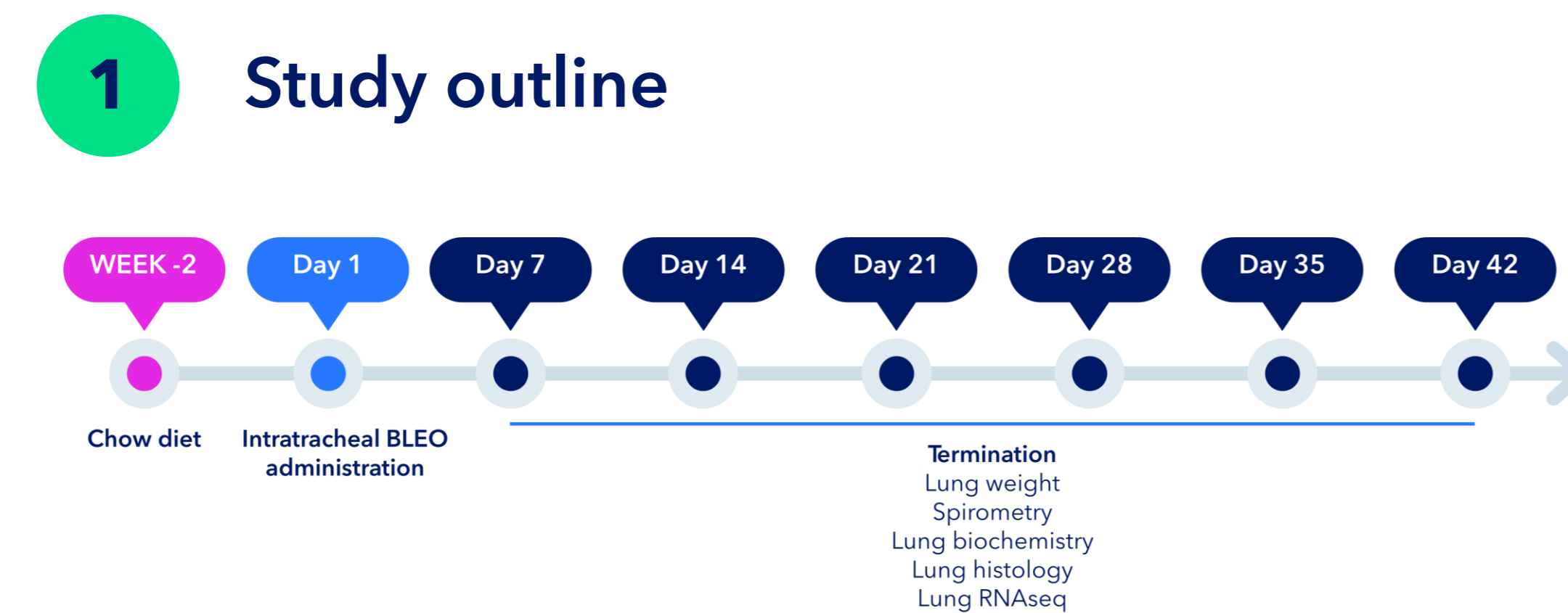
10-12 weeks old C57BL/6J male mice received either a single intratracheal instillation of bleomycin (1.5 mg/kg, 50 µL) or saline (CTRL) at study day 1. BLEO-IPF animals were randomized into study groups based on body weight loss at day 6 post-BLEO and terminated at specified time points (Fig. 1).

Terminal pulmonary endpoints included (1) spirometry (flexiVent) for expiratory/inspiratory capacity; (2) biochemical analysis for hydroxyproline (HP) content; (3) quantitative histomorphometry for markers of cellular senescence (p21) and fibrosis (PSR, Col1a1); (4) AI-based Ashcroft fibrosis scoring; (5) transcriptome signatures of cellular senescence.



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Group	Animal	Name	Number of animals
1	CTRL	CTRL	10
2	BLEO-IPF	D7	10
3	BLEO-IPF	D14	11
4	BLEO-IPF	D21	12
5	BLEO-IPF	D28	12
6	BLEO-IPF	D35	12
7	BLEO-IPF	D42	9

Figure 1. Study outline and group overview

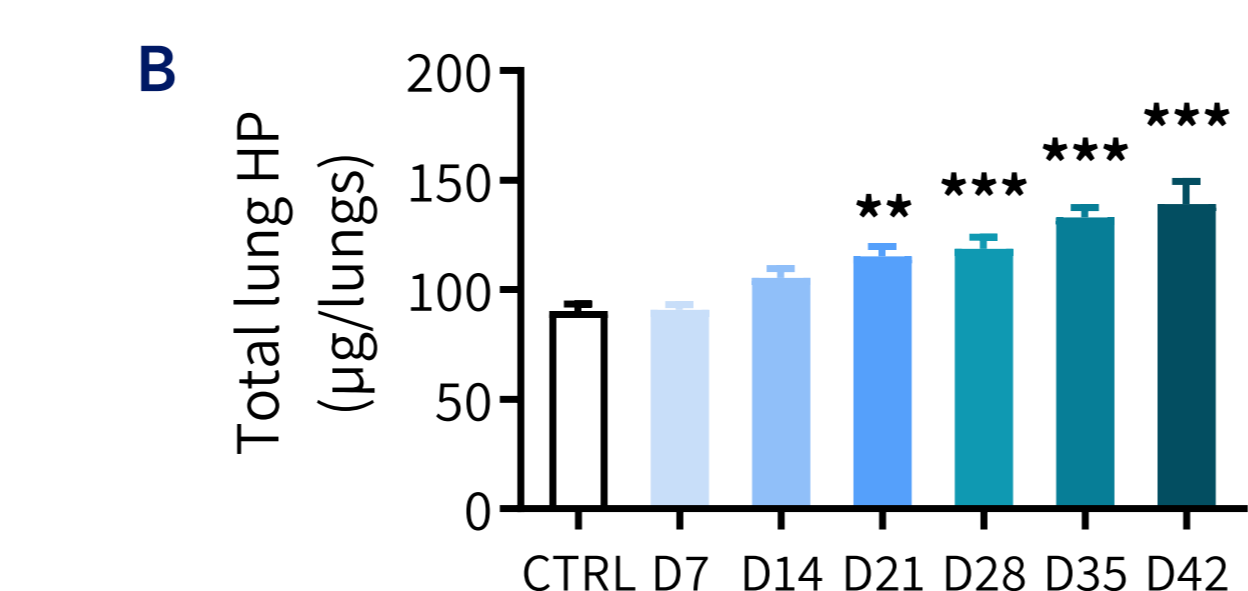
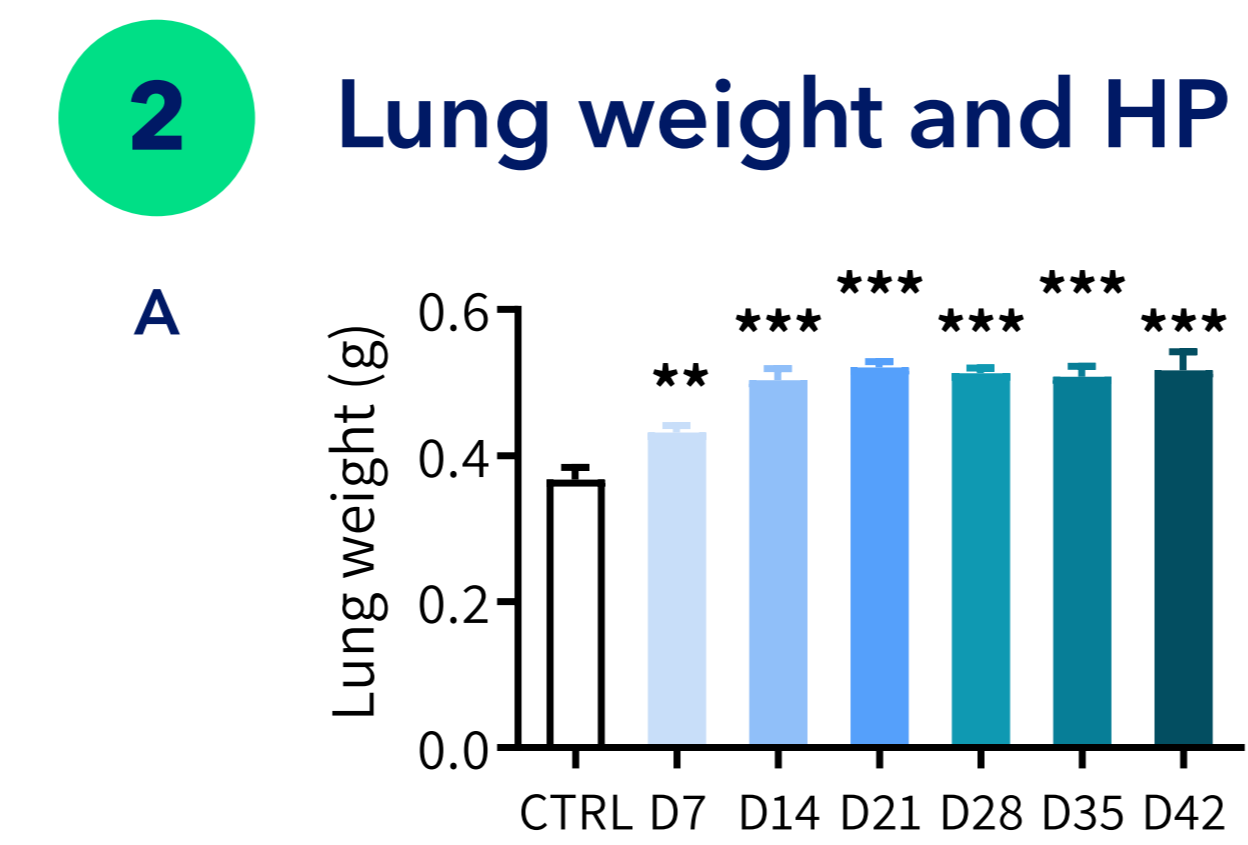


Figure 2. Metabolic and biochemical parameters in BLEO-IPF mice. (A) Terminal lung weight (g). (B) Terminal lung total HP. Dunnett's test one-factor linear model. **p<0.01, and ***p<0.001 vs. CTRL group.

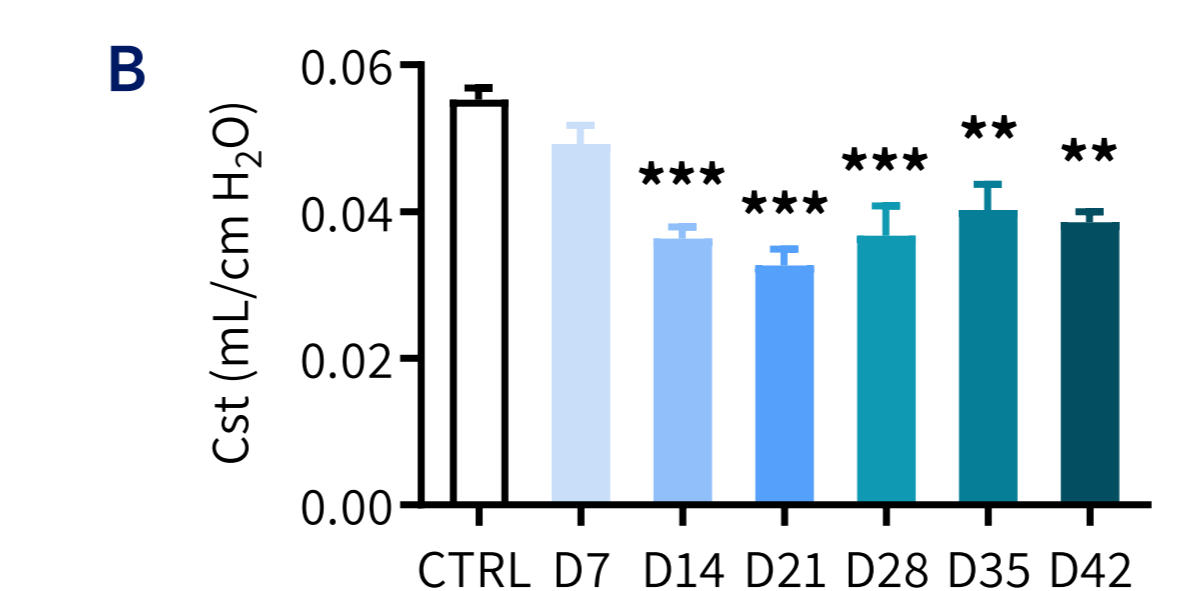
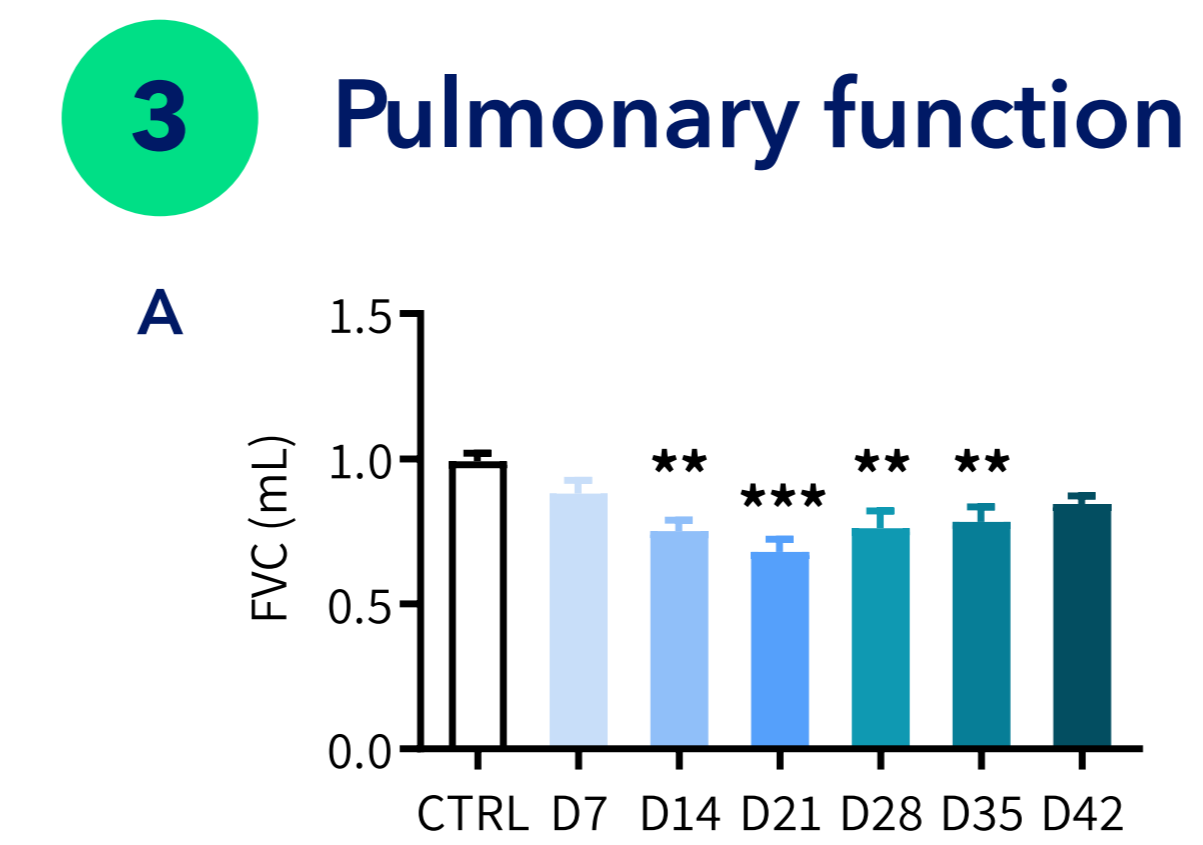


Figure 3. Pulmonary function testing in BLEO-IPF mice. (A) Forced vital capacity (FVC). (B) Static compliance (Cst). Dunnett's test one-factor linear model. **p<0.01, and ***p<0.001 vs. CTRL group.

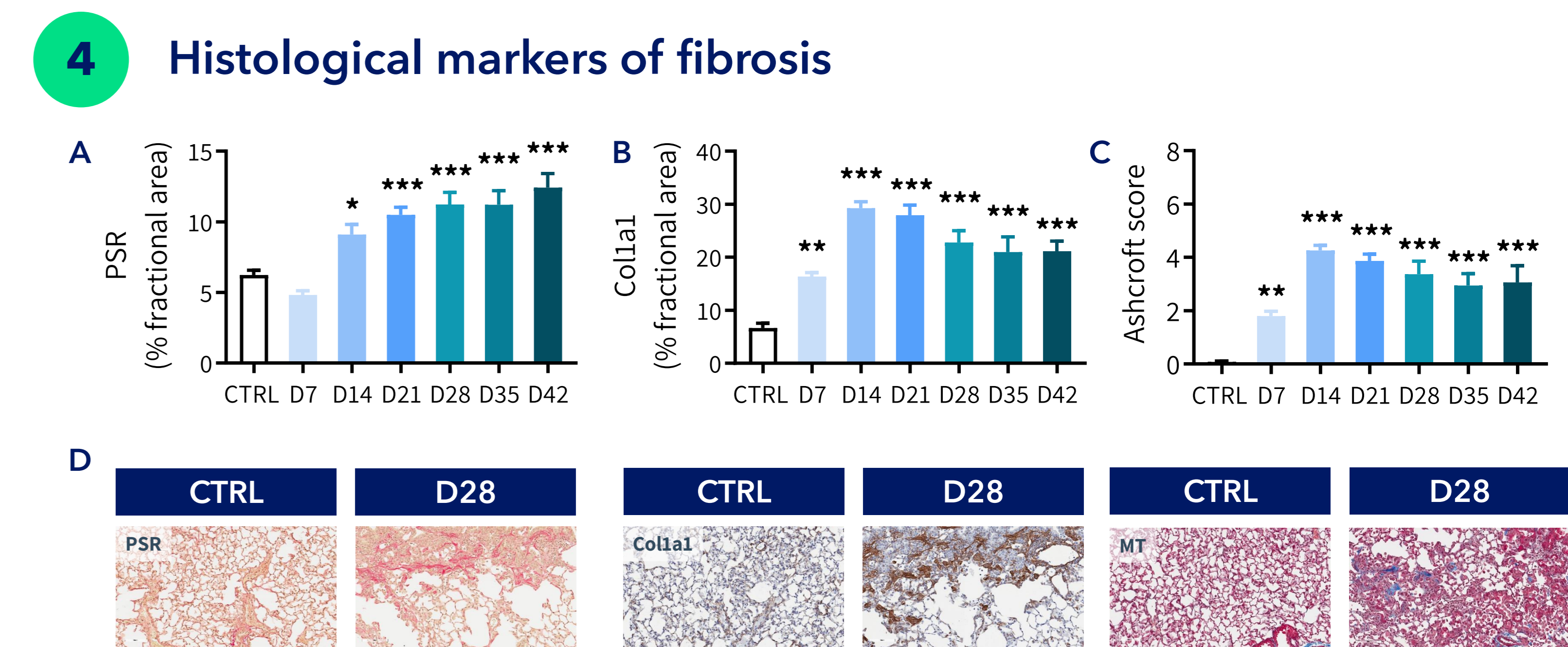


Figure 4. Lung quantitative histological markers of fibrosis. Histomorphometric assessments of PSR and Col1a1 were performed by conventional image analysis. Data were calculated as proportionate (% area) of histological staining (mean ± SEM). (A) % fractional area of PSR-stained fibers; (B) % fractional area of Collagen-1α1. Histopathological Ashcroft score were determined by GHOST deep learning-based image analysis. (C) Ashcroft score by Gubra Histopathological Objective Scoring Technique (GHOST). Dunnett's test one-factor linear model. *p<0.05, **p<0.01 and ***p<0.001 vs. CTRL group. (D) Representative photomicrographs of lung histological stainings.

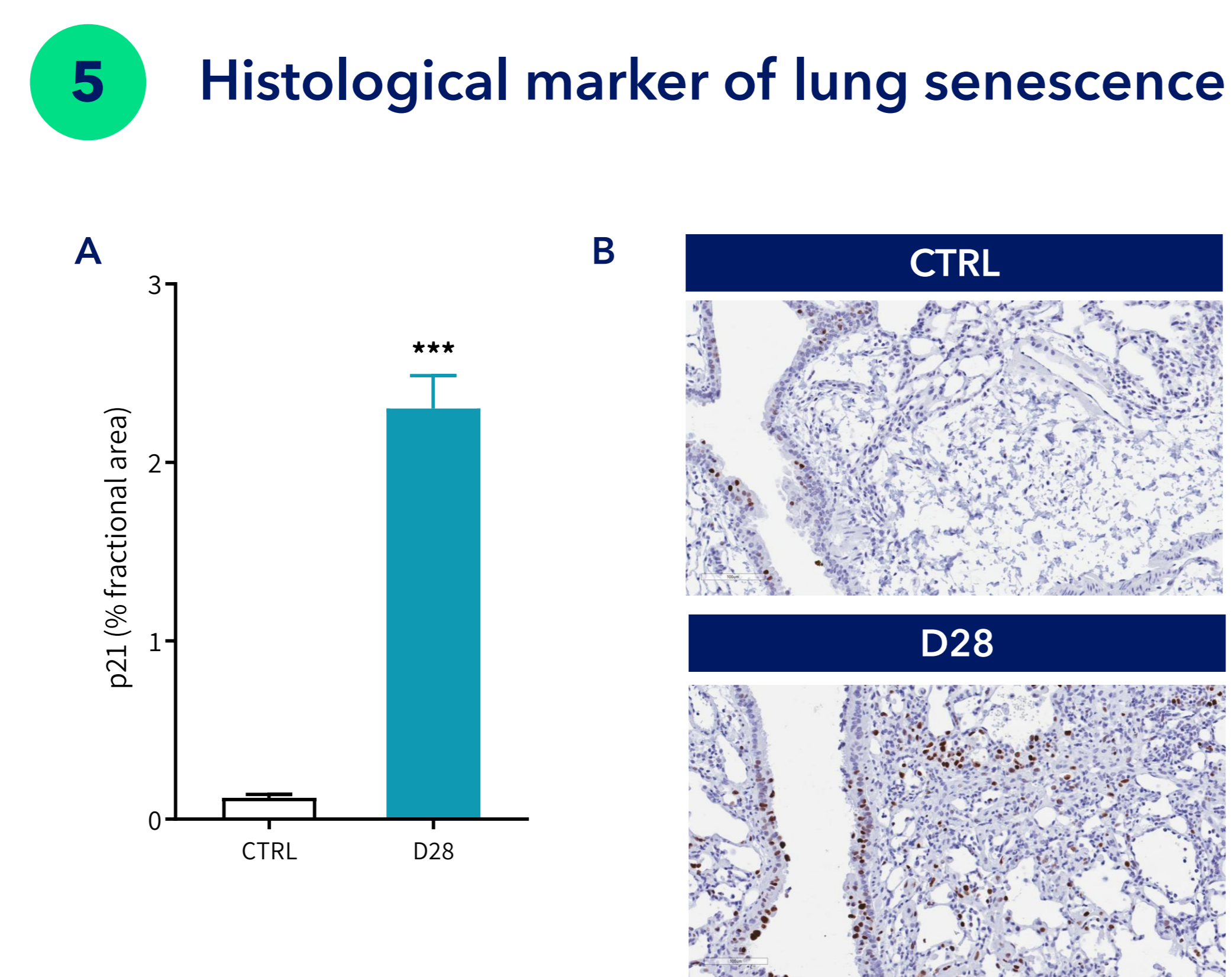


Figure 5. Lung quantitative histological marker of cellular senescence. Histomorphometric assessment of p21 expression was performed by conventional image analysis. Data were calculated as proportionate (% area) of histological staining (mean ± SEM). (A) % fractional area of p21. (B) Representative photomicrographs of lung histological stainings. Dunnett's test one-factor linear model. ***p<0.001 vs. CTRL group.

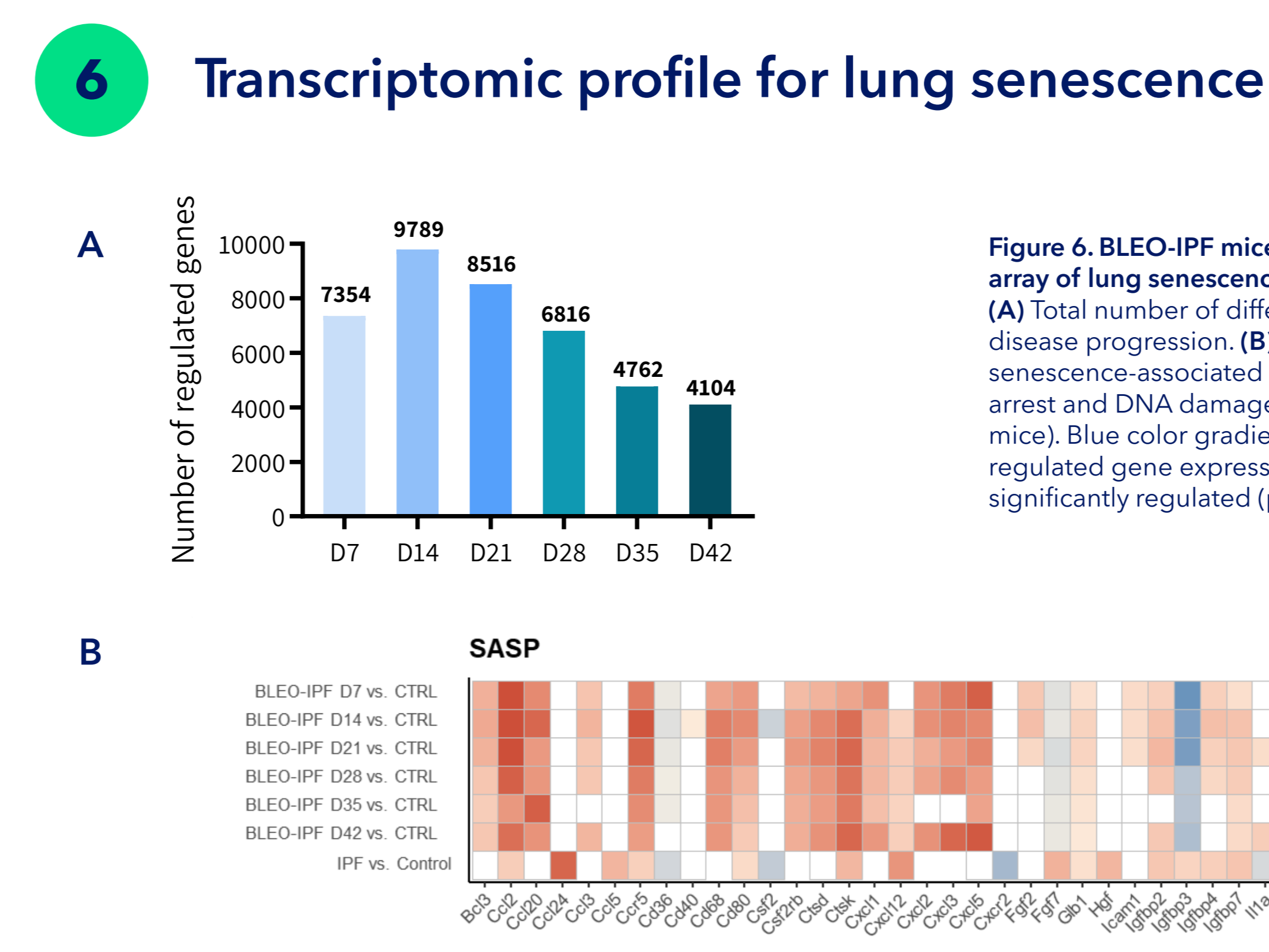


Figure 6. BLEO-IPF mice show marked upregulation of a wide array of lung senescence-associated genes. (A) Total number of differentially expressed genes (DEGs) during disease progression. (B) Candidate genes associated with the senescence-associated secretory phenotype (SASP), cell cycle arrest and DNA damage (log₂-fold change compared to CTRL mice). Blue color gradients indicate significantly (p<0.05) down-regulated gene expression. White boxes indicate genes not significantly regulated (p>0.05) compared to CTRL group.

Conclusion

- + BLEO-IPF mice demonstrate progressive increase in lung weight and total hydroxyproline content
- + BLEO-IPF mice demonstrate reduced pulmonary expiratory and inspiratory function.
- + BLEO-IPF mice demonstrate increased lung levels of quantitative histological markers of fibrosis
- + BLEO-IPF mice demonstrate increased lung level of p21, a well-established histological marker of cellular senescence
- + BLEO-IPF mice demonstrate marked lung transcriptome perturbations, including upregulation of a wide array of cellular senescence-associated genes

The single-installation BLEO-IPF mouse model represents a translational preclinical model for profiling potential senotherapeutics for IPF